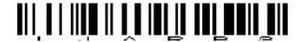

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Review Article



Molecular mechanism and systemic response of erythropoietin:A Review

Obeagu,Emmanuel Ifeanyi^{1*},Okoroiwu,I.L.²,Obeagu,GetrudeUzoma³

1.Diagnostic Laboratory Unit,Department of University Health Services,Michael Okpara University of Agriculture,UmuDike,Abia State,Nigeria.

2.Department of Medical Laboratory Science,Imo State University,Owerri,Nigeria.

3.Departemnt of Nursing Science,Ebonyi State University,Abakaliki,Nigeria.

Corresponding Authour Address: Diagnostic Laboratory Unit,Department of University Health Services,Michael Okpara University of Agriculture,UmuDike,Abia State,Nigeria.

*Corresponding author

Abstract

This paper reviews the molecular mechanism and systemic response of erythropoietin (Epo).Erythropoietin(Epo) is a glycoprotein hormone that causes the production of red blood cell.This helps to increase the level of haemoglobin and packed cell volume.It is produced mainly in the interstitial fibroblast of the renal cortex.It has additional nonhaematopoietic effects.Erythropoietin enhancer is activated by hypoxia-inducible transcription factors (HIFs) which is composed of an O₂-labile -subunit and a constitutive subunit.Erythropoietin synthesis follows a negative feedback mechanism and is regulated by hypoxia.There is an inverse relationship between erythropoietin (Epo) level and haematocrit (Hct) or haemaoglobin (Hb) concentration.The level of erythropoietin varies greatly between individuals and there is no serious sex or age differences.This diurnal fluctuation with a nadir occuring in the morning.Therefore,the kidneys should be kept in good health conditions always to maintain the daily requirement of erythropoietin level to prevent anaemia and apoptosis of the red blood cells.

Keywords:Erythropoietin,Molecular Mechanism , Systemic Response Of Erythropoietin,Regulation Of Erythropoietin Synthesis

Introduction

The hormone erythropoietin (Epo) maintains red blood cell mass by promoting the survival,proliferation and differentiation of erythrocytic progenitors (Jelkman,2011).Erythropoietin is essential for erythropoiesis.Erythropoietin is a 30.4Kda glycoprotein produced by the kidney,and is mostly well-known for its physiological function in regulating red blood cell production in the bone marrow (Moore and Bellomo,2011).Erythropoietin has additional protective effects,which may be useful in the prevention or treatemnt of acute kidney injury.These protective mechanisms are multifactorial in nature and include inhibition of apoptotic cell death,stimulation of cellular regeneration,inhibition of deleterious pathways, and promotion of recovery (Moore and

Belloman,2011).Erythropoietin originates mainly from fibroblasts in the renal cortex.

Physiology of erythropoietin

Erythropoietin is primarily expressed by hepatocytes during the fetal state.After birth,peritubular fibroblasts in the renal cortex become the main production site.Erythropoietin synthesis is controlled at the transcriptional level.Erythropoietin mRNA is also detectable in brain,liver,spleen,lung and testis,but these organs are not able to substitute for renal erythropoietin in chronic kidney disease (CKD).Brain-derived erythropoietin acts locally as a neuroprotective factor (Noguchi et al.,2007).Erythropoietin is an acidic

glycoprotein of about 30.4 Kda and comprises of 165 amino acids and four glycans. Circulating erythropoietin exhibits several glycosylation isoforms that differ in electrical charge and biological activity. Erythropoietin amounts are usually expressed in international units (IU), with one IU exerting the same erythropoiesis stimulating activity in rodents as 5 μ mol cobaltous chloride (Jelkman, 2007). Systemic erythropoietin is an anti-apoptotic agent for erythrocytic progenitors, predominantly the colony-forming units-erythroids (CFU-Es). In response to erythropoietin the CFU-Es proliferate and differentiate to generate cohorts of proerythroblasts and normoblasts. The human haematopoietic Epo-receptor (Epo-R) is a 484 amino acid glycoprotein of about 60 Kda, which belongs to the cytokine class I receptor family and forms homodimers. On the binding of erythropoietin to the Epo-R dimer, cytoplasmic Janus Kinases 2 (JAK2) catalyse the phosphorylation of tyrosine residues of the Epo-R and of various intracellular proteins (Jelkman, 2011).

A role has been proposed for erythropoietin as a cytoprotective agent for several non-haematopoietic tissues, including the brain, the heart, blood vessels and the kidneys (Brines and Cerami, 2006). Some researchers believe that the effects of erythropoietin on non-erythrocytic cells are mediated by a heterodimeric receptor consisting of one Epo-R molecule in complex with a dimer of the common cytokine receptor (CR). The Epo-R/CR concept is mainly based on the observation that erythropoietically inactive erythropoietin derivatives and analogues provide tissue protection in animal models (Leist et al., 2004).

Molecular mechanism of erythropoietin synthesis by hypoxia

The mechanisms of the renal and hepatic erythropoietin expression differ. Renal cells respond in an all-or-nothing fashion to hypoxia (Koury et al., 1989), whereas hepatoma cells respond in a graded way. The hypoxia-response elements (HREs) in control of the erythropoietin gene are located upstream in the kidney but downstream in the liver as opined by Kochling et al., 1998). The erythropoietin promoter is suppressed by GATA-2 in normoxia (Tsuchiya et al., 1997). GATA-2 levels decrease in hypoxia (Imagawa et al., 2003). Erythropoietin enhancer is activated by hypoxia-inducible transcription factors (HIFs). These are composed of an O₂-labile

-subunit (120KDa; Isoforms 1, 2 or 3) and a constitutive β -subunit (90-95Kda) (Warnecke et al., 2004; Haase, 2010). HIF-2 is activated by the stress-responsive deacetylase sirtuin 1 (Dioum et al., 2009). Interestingly, hepatocyte-derived HIF-2 is also involved in the regulation of iron metabolism genes, supporting a role for HIF-2 in the coordination of erythropoietin synthesis with iron homeostasis (Kapitsinou et al., 2010). HIFs cooperate with hepatocyte nuclear factor 4 (HNF-4) and the transcriptional co-activators P300 and cAMP response element-binding protein (CREB)-binding protein (CBP) (Bunn et al., 1998).

The C-terminus of the HIF- β subunits comprises O₂-dependent degradation domains (O-DDD) that are properly hydroxylated in the presence of O₂ (Epstein et al., 2001; Ivan et al., 2001; Yu et al., 2001; Jacakkola et al., 2001; Bruick and McKnight, 2001). The reaction is catalysed by specific Fe²⁺-containing prolyl-4-hydroxylase (PHD-1, -2, and -3), which transfer one O-atom to the proline and the other to 2-oxoglutarate yielding CO₂ and succinate (Bruegge et al., 2007). The properly hydroxylated HIF- β combines with Von Hippel-Lindau tumour suppressor protein (VHL)/E3 ligase and promptly undergoes proteosomal degradation (Huang et al., 1998; Maxwell et al., 1999). As PHD-2 and PHD-3 are themselves HIF-target genes, their expression increases and HIF- β levels decline during long-term hypoxic periods (del Peso et al., 2003; Marxsen et al., 2004; Aprelikova et al., 2004; Stiehl et al., 2006). The transcriptional activity of the HIFs is suppressed by HIF- β asparinylation, which prevents the binding of the transcriptional co-activator CBP/P300. The reaction is catalysed by the factor inhibiting HIF-1 (Fahnestock et al., 2001; McNeill et al., 2002), another Fe²⁺ is required for HIF- β degradation may provide an explanation for the increased plasma Epo level in patients treated with the iron chelator deferoxamine (Kling et al., 1996). Cobalt is thought to enhance Epo expression by replacing the essential Fe²⁺ in the HIF- β dioxygenases, which results in HIF- β stabilisation. 2-oxoglutarate competitors likewise inhibit the hydroxylation of HIF- β .

Systemic erythropoietin response

The major roles of erythropoietin and hemoglobin are to keep red blood cell mass and hemoglobin constant day by day, and to hasten red blood cell recovery after

haemorrhage. The basal plasma concentration of Epo ranges from 6 to 32 iu/l. The levels vary greatly between individuals, with the result that significant sex or age-specific differences cannot be detected. Of note is the diurnal fluctuation, with a nadir in the morning. The response is dynamic, with initially very high Epo values that drop towards the normal ones before haemoglobin normalises. The mechanism of rapid decrease may in part be caused by lowered HIF-levels during long-term hypoxia (Stiehl et al., 2006). Epo level is not only dependent on the rate of Epo production, but also on its removal. In vitro studies have demonstrated that Epo is internalised and degraded by its target cells (Gross and Lodish, 2006). Anaemic patients with bone marrow hypoxia exhibit extremely high plasma Epo level (10,000 iu/l or more) compared with subjects suffering from haemolytic anaemia.

Because Epo production depends on the tissue PO₂ declines or when the O₂ affinity of the blood increases. On ascent to altitude, Epo levels reach peak values after 1-2 days and then fall to a new plateau at about twice that present at sea level (Abbrecht and Littell, 1972).

Role of angiotensin II

The signal to produce more erythrocytes following haemorrhage is apparently linked to the signals to retain salt and water by means of the renin-angiotensin system (Dunn et al., 2007). Angiotensin II (Ang II) is thought to stimulate erythropoiesis by two means. First, Ang II increases Epo production. Second, Ang II is a growth factor for the myeloid erythrocytic progenitors (Dunn et al., 2007; Vlahakos et al., 2010). The infusion of Ang II has been shown to raise the plasma Epo level to 17 iu/l in healthy men (Glossman et al., 2001).

A feedback regulation of red cell mass and blood volume by means of Epo and Ang II seems to exist. Treatment of healthy men with recombinant human Epo (rhEpo) produces an increase in red cell mass. However, the increase in haematocrit (Hct) is accompanied by a decrease in plasma volume, which is probably due to a down-regulation of the renin-angiotensin-aldosterone system and results in a constancy of the blood volume. Thus, Epo treatment in healthy humans raises haemoglobin by two

mechanisms: an increase in red cell mass, and a decrease in plasma volume (Lundby et al., 2007).

Conclusion

Erythropoietin (Epo) is a glycoprotein hormone that regulates erythropoiesis and other nonhaematopoietic functions. It is mainly produced in the interstitial fibroblasts of renal cortex in adults and in the liver cells in the foetus. It performs its functions by binding to erythropoietin receptor (Epo-R) which induces the expression of hypoxia-inducible factor (HIF) which regulates the synthesis of erythrocytes. Erythropoietin can be used in the treatment of anaemia and will reduce the rate of blood transfusion among patients. The kidneys should be properly cared to ensure the continual sustenance of the tissues where the erythropoietin for the upkeep of the human system.

References

- Abbrecht, P.H. and Littell, J.K. (1972). Plasma Erythropoietin in men and mice during acclimatisation to different altitudes. *J. Appl. Physiol.* 32:54-58.
- Aprelikova, O., Chandramouli, G.V., Wood, M., Vasselli, J.R., Riss, J., Maranchie, J.K., Linehan, W.M. and Barret, J.C. (2004). Regulation of HIF prolyl hydroxylases by hypoxia-inducible factors. *J. Cell Biochem* 92:491-501.
- Brines, M. and Cerami, A. (2006). Discovering erythropoietin's extra-haematopoietic functions: biology and clinical promise. *Kidney Int* 70:246-250.
- Bruegge, K., Jelkman, W. and Metzner, E. (2007). Hydroxylation of hypoxia-inducible transcription factors and chemical compounds targeting the HIF-hydroxylases. *Curr. Med. Chem* 14:1853-1862.
- Bruick, R.K. and McKnight, S.L. (2001). A conserved family of prolyl-4-hydroxylases that modify HIF. *Science* 294:1337-1340.
- Bunn, H.F., Gu, J., Huang, L.E., Park, J.W. and Zhu, H. (1998). Erythropoietin: a model system for studying oxygen-dependent gene regulation. *J. Exp. Biol.* 201:1197-1201.
- del Peso, L., Castellanos, M.C., Temes, E., Martin-Puig, S., Cuevas, Y., Olmos, G. and Landazuri, M.O. (2003). The Von Hippel Lindau/hypoxia inducible factor (HIF) pathway regulates the

- transcription of the HIF-proline hydroxylase genes in response to low oxygen. *J. Biol. Chem.* 278:48690-48695.
- Dioum, E.M., Chen, R., Alexander, M.S., Zhang, Q., Hogg, R.T., Gerard, R.D. and Garcia, J.A. (2009). Regulation of hypoxia-inducible factor 2 signaling by the stress-responsive deacetylase sirtuin 1. *Science* 324:1289-1293.
- Dunn, A., Lo, V. and Donnelly, S. (2007). The role of kidney in blood volume regulation: the kidney as a regulator of the haematocrit. *Am. J. Med. Sci.* 334:65-71.
- Epstein, A.C.R., Gleadle, J.M., McNeill, L.A., Hewitson, K.S., O'Rourke, J., Mole, D.R., Mukherji, M., Metzger, E., Wilson, M.I., Dhanda, A., Tian, Y.M., Masson, N., Hamilton, D.L., Jaakkola, P., Barstead, R., Hodgkin, J., Maxwell, P.H., Pugh, C.W., Schofield, C.J. and Ratcliffe, P.J. (2001). C.elegans EGL-9 and mammalian homologs define a family of dioxygenases that regulate HIF by prolyl hydroxylation. *Cell* 107:43-54.
- Gossman, J., Burkhardt, R., Harder, S., Lenz, T., Sedlmeyer, A., Klinkhardt, U., Geiger, H. and Scheuermann, E.H. (2001). Angiotensin II infusion increases plasma erythropoietin levels via an angiotensin II type I receptor-dependent pathway. *Kidney Int.* 60:83-86.
- Gross, A.W. and Lodish, H.F. (2006). Cellular trafficking and degradation of erythropoietin and novel erythropoiesis stimulating protein (NESP). *J. Biol. Chem.* 281:2024-2032.
- Haase, V.H. (2010). Hypoxic regulation of erythropoiesis and iron metabolism. *Am. J. Physiol. Renal Physiol.* 299:F1-F13.
- Huang, L.E., Gu, J., Schau, M. and Bunn, H.F. (1998). Regulation of hypoxia-inducible factor 1 is mediated by an O₂-dependent degradation domain via the ubiquitin-proteasome pathway. *Proc. Natl. Acad. Sci. U.S.A.* 95:7987-7992.
- Imagawa, S., Nakano, Y., Obara, N., Suzuki, N., Doi, T., Kodama, T., Nagasawa, T. and Yamamoto, M. (2003). A GATA-specific inhibitor (K-7174) rescues anaemia induced by IL-1, TNF- α , or 1-NMMA. *FASEB J.* 17:1742-1744.
- Ivan, M., Kondo, K., Yin, H., Kim, W., Valiando, J., Ohh, M., Salic, A., Asara, J.M., Lane, W.S., and Kaelin, W.G.J. (2001). HIF targeted for VHL-mediated destruction by proline hydroxylation: implications for O₂ SENSING. *Science* 292:464-468.
- Jaakkola, P., Mole, D.R., Tian, Y.M., Wilson, M.I., Gielbert, J., Gaskell, S.J., Kriegsheim, A., Hebestreit, H.F., Mukherji, M., Schofield, C.J., Maxwell, P.H., Pugh, C.W. and Ratcliffe, P.J. (2001). Targeting of HIF-1 to Von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science* 292:468-472.
- Jelkman, W. (2007). Erythropoietin after a century of research: younger than ever. *Eur. J. Haematol.* 78:183-205.
- Jelkman, W. (2011). Topical review: Regulation of erythropoietin production. *J. Physiol.* 589(6):1251-1258.
- Kapitsinou, P.P., Liu, Q., Unger, T.L., Rha, J., Davidoff, O., Keith, B., Epstein, J.A., Moores, S.L., Erickson-Miller, C.L. and Haase, V.H. (2010). Hepatic HIF-2 regulates erythropoietic responses to hypoxia in renal anaemia. *Blood* 116:3039-33048.
- Kling, P.J., Drastin, P.R., Roberts, R.A., Dos Santos, B., Brooks, D.L., Hedlund, B.E. and Taetle, R. (1996). Iron deprivation increases erythropoietin production in vitro, in normal subjects and patients with malignancy. *Br. J. Haematol.* 95:241-248.
- Kochling, J., Curtin, P.T. and Madan, A. (1998). Regulation of human erythropoietin gene induction by upstream flanking sequences in transgenic mice. *Br. J. Haematol.* 103:960-968.
- Koury, S.T., Koury, M.L., Bondurant, M.C., Caro, J. and Graber, S.E. (1989). Quantitation of erythropoietin-producing cells in kidneys of mice by in situ hybridisation: correlation with haematocrit, renal erythropoietin mRNA, and serum erythropoietin concentration. *Blood* 74:645-651.
- Leist, M., Ghezzi, P., Grasso, G., Biancho, R., Villa, P., Fratelli, M., Savino, C., Bianchi, M., Nielson, J., Gerwien, J., Kallunki, P., Larsen, A.K., Helboe, L., Christensen, S., Pedersen, L.O., Nielson, M., Torup, L., Sager, T., Sforza, A., Erbayraktar, S., Erbayraktar, Z., Gokmen, N., Yilmaz, O., Cerami-Hand, C., Xie, Q.W., Coleman, T., Cerami, A. and Brines, M. (2004). Derivatives of erythropoietin that are tissue protective but not erythropoietic. *Science* 305:239-242.
- Lundby, C., Thompson, J.J., Boushel, R., Koskolou, M., Warberg, J., Calbet, J.A. and Robach, P. (2007). Erythropoietin treatment elevates haemoglobin concentration by increasing red cell volume and depressing plasma volume. *J. Physiol.* 578:309-314.
- McNeill, L.A., Hewitson, K.S., Klaridge, T.D., Seibel, J.F., Horsefall, L.E. and Schofield, C.J. (2002). Hypoxia-inducible factor asparaginyl hydroxylase (FIH-1)

- catalyses hydroxylation at the beta-carbon of asparagine-803. *Biochem J.* 367:571-575.
- Mahon, P.C., Hirota, K. And Semenza, G.L. (2001). FIH-1: a novel protein that interacts with HIF-1 and VHL to mediate repression of HIF-1 transcriptional activity. *Genes Dev.* 15:2675-2686.
- Marxsen, J.H., Stengel, P., Doege, K., Heikkinen, P., Jokilehto, T., Wagner, T., Jelkman, W., Jaakkola, P. and Metzen, E. (2004). Hypoxia-inducible factor-1 promotes its degradation by induction of HIF-1 prolyl-4-hydroxylases. *Biochem J.* 381:761-767.
- Maxwell, P.H., Wiesener, M.S., Change, G.W., Clifford, S.C., Vaux, E.C., Cockman, M.E., Wykoff, C.C., Pugh, C.W., Maher, E.R. and Ratcliffe, P.J. (1999). The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature* 399:271-275.
- Moore, E. and Bellomo, R. (2011). Erythropoietin (Epo) in acute kidney injury. *Ann. Intensive Care* 1:3
- Noguchi, C.T., Asavaritikrai, P., Teng, R. and Jia, Y. (2007). Role of erythropoietin in the brain. *Crit. Rev. Oncol. Haematol.* 64:159-171.
- Stiehl, D.P., Wirthner, R., Koditz, J., Spielmann, P., Camenisch, G. and Wenger, R.H. (2006). Increased prolyl 4-hydroxylase domain proteins compensate for decreased oxygen levels. Evidence for an autoregulatory oxygen-sensing system. *J. Biol Chem* 281:23482-23491.
- Tsuchiya, T., Okada, M., Ueda, M. and Yasukochi, Y. (1997). Activation of the erythropoietin promoter by a point mutation of the GATA to TATA in the -30 region. *J. Biochem* 121:193-196.
- Vlahakos, D.V., Marathias, K.P., and Madias, N.E. (2010). The role of the renin-angiotensin system in the regulation of erythropoiesis. *Am. J. Kidney Dis.* 56:558-565.
- Warnecke, C., Zaborowska, Z., Kurreck, J., Erdmann, V.A., Fret, U., Wiesener, M. and Eckardt, K.U. (2004). Differentiating the functional role of hypoxia-inducible factor (HIF)-1 and HIF-2 (EPAS-1) by the use of RNA interference: erythropoietin is a HIF-2 target gene in Hep3B and Kelly cells. *FASEB* 18:1462-1464.
- Yu, F., White, S.B., Zhao, Q. and Lee, F.S. (2001). HIF-1 binding to VHL is regulated by stimulus-sensitive proline hydroxylation. *Proc. Natl. Acad. Sci. U.S.A.* 98:9630-9635.